



# Comparing Selection Coefficients and Omega for Codon Substitution Rates

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# Agenda

- Introduction
- Mutation Selection Model
  - Mutation Rate
  - Selection on Codon Usage
  - Selection on Protein
- Omega Model
  - Fixed Effects Likelihood
  - Random Effects Likelihood
- Omega-MutSel Comparison
- Research Goals

# Introduction

- Codon Bias
  - Different frequencies for synonymous codons that code for the same amino acid
  - There is some external selective pressure
- How do we infer positive selection from DNA?

Answer: Two models to examine selection at individual sites

  1. Selection Coefficient
  2. Omega

# Motivation

- Everyone uses the Omega model
  - Easy to run
  - Been around for a long time
  - However, it leaves out information about underlying evolution
- Recently, mutation selection models have been developed to model selection pressure
- We don't know which model is better

# Software Packages

- Phylogenetic Analysis by Maximum Likelihood (PAML)
  - Estimates selective strengths on codon usage
- Hypothesis testing using Phylogenies (HyPhy)
- Software listed above can take a very long time (months)
- Need to get accustomed to software – what output, what parameters are required as input, etc.

# Mutation Selection Model<sup>1</sup> (MutSel)

- Looks at the balance of mutation and selection
- Assume only one nucleotide change at a time
- Models the following:
  1. Nucleotide Mutation
  2. Selection on Codon Usage
  3. Selection on the Protein

[1] Yang, Z., and R. Nielsen. 2008. Mutation-Selection Models of Codon Substitution and Their Use to Estimate Selective Strengths on Codon Usage.

# Mutation Rate

First we need to define some variables:

$\mu_{ij}$  = mutation rate of nucleotide  $i$  to  $j$  in one generation

$a_{ij}$  = nucleotide substitution rate from  $i$  to  $j$  from GTR<sup>1</sup> matrix

$\pi_j^*$  = **mutation bias**; we scale  $\pi_j^*$  such that  $\sum \pi_j^* = 1$

Now we can calculate the mutation rate:

$\mu_{ij} = a_{ij}\pi_j^*$  where  $a_{ij} = a_{ji}$  for all  $i \neq j$

[1] Tavaré, S. 1986. "Some Probabilistic and Statistical Problems in the Analysis of DNA Sequences". Lectures on Mathematics in the Life Sciences (American Mathematical Society) 17: 57–86.

# Selection on Codon Usage

Definitions:

$f_I$  = fitness parameter for codon  $I$

$s_{IJ} = f_J - f_I =$  **selection coefficient** for the mutation that changes codon  $I$  into  $J$

To calculate the fixation probabilities:

$S_{IJ} = 2Ns_{IJ} = 2N(f_J - f_I) =$  **scaled selection coefficient**

$N$  = population size

$h(S_{IJ}) = S_{IJ}/(1 - e^{-S_{IJ}}) =$  **ratio** of fixation probability of the  $I \rightarrow J$  mutation to the fixation probability of a neutral mutation



# Selection on Codon Usage

- Let  $Q$  denote the codon substitution matrix:

$$q_{IJ} = \begin{cases} 0 & \text{if more than one change} \\ \mu_{i_k j_k} h(S_{IJ}) & \text{if synonymous substitution} \\ \omega \mu_{i_k j_k} h(S_{IJ}) & \text{if non-synonymous substitution} \end{cases}$$

Where  $k$  is the codon position in the sequence

- Why use  $\omega$ ?
  - Because it is simple and it produces similar estimates of mutation parameters as models that incorporate chemical properties<sup>1</sup>

[1] Yang, Z., and R. Nielsen. 2008. Mutation-Selection Models of Codon Substitution and Their Use to Estimate Selective Strengths on Codon Usage.

# Selection on the Protein

- Averaged over time, the proportion of  $I \rightarrow J$  mutations among all mutations is:

$$m_{IJ} = \frac{\pi_I \mu_{i_k j_k}}{\sum_{I \neq J} \pi_I \mu_{i_k j_k}} \quad \text{and} \quad m_{IJ}^+ = \frac{\pi_I \mu_{i_k j_k} \mathbb{1}}{\sum_{I \neq J} (\pi_I \mu_{i_k j_k} \mathbb{1})}$$

Note:  $\mu_{ij} = a_{ij} \pi_j^*$  = mutation rate;  $S_{IJ}$  = scaled selection coefficient

- Where  $\mathbb{1}$  is the indicator function:
  - $\mathbb{1} = 1$  if  $S_{IJ} > 0$ , and 0 otherwise
  - Only include advantageous mutations
- Thus, the strength of **positive selection** on the protein is:

$$\bar{S}_+ = \sum_{I \neq J} (m_{IJ}^+ S_{IJ} \mathbb{1})$$

# Omega Models

- Compare synonymous and non-synonymous mutations

- $\omega = \frac{dN}{dS}$

- if*  $\omega < 1$  implies purifying selection

- $\omega = 1$  implies neutral mutations

- $\omega > 1$  implies diversifying positive selection

- Typically calculated by taking average over all codons
- Problem: It becomes difficult for  $\omega > 1$
- Possible Solution: Create statistical models for  $\omega$

[1] Z. Yang, et al. 2000. Codon-Substitution Models for Heterogeneous Selection Pressure at Amino Acid Sites.

# Random Effects Likelihood (REL)

- If we use one  $\omega$  for each site, we get too many parameters
- Probability of observing data  $x_h$  given site  $h$ :

$$f(x_h) = \sum_{k=1}^2 p_k f(x_h | \omega_k) = p_1 f(x_h | \omega_1) + p_2 f(x_h | \omega_2)$$

$h = \{1, 2, \dots, n\}$  and  $p$  = proportion of codon sites in categories

Two categories:

1. Non-synonymous mutations are neutral
2. Non-synonymous sites are eliminated by selection

R. Nielsen and Z. Yang. 1998. Likelihood Models for Detecting Positively Selected Amino Acid Sites and Applications to the HIV-1 Envelope Gene.

# Fixed Effects Likelihood (FEL)

- Keep the model parameters fixed:
  - Branch lengths
  - Nucleotide rate biases
  - Tree topology
- Using a FEL rate matrix<sup>1</sup>, we can compute each site
- Apply a likelihood test to determine significance
  
- Can process gene-size alignments of several hundred sequences in a few hours on a small cluster

[1] S. Pond, and S. Frost. 2005. Not So Different After All: A comparison of Methods for Detecting Amino Acid Sites Under Selection

# Omega-MutSel Comparison

$\lambda_a$  = parameter determining frequency of amino acid A  
“scaled selection coefficient”

$$F(a) \sim e^{-\lambda \text{int}(aa)} = \text{fitness}$$

$$\pi_{a \rightarrow b} = \frac{1 - [F(a)/F(b)]^{\frac{1}{N}}}{1 - F(a)/F(b)}$$

$$K = \mu N \sum_a [F(a) \sum_b \pi_{a \rightarrow b}]$$

- If we perform some algebra on  $\pi_{a \rightarrow b}$ , we can eliminate  $N$  from the K equation.

[1] D. Ramsey, M. Scherrer, T. Zhou, and C. Wilke. The Relationship Between Relative Solvent Accessibility and Evolutionary Rate in Protein Evolution. *Genetics* 2011.

# Omega-MutSel Comparison

$$dN = \frac{K_N}{N_N} = \frac{\mu N \sum_i \sum_{j \in \mathcal{M}} F(i) \pi_{i \rightarrow j}}{\sum_i \sum_{j \in \mathcal{M}} F(i)}$$

$$dS = \frac{K_S}{N_S} = \mu$$

- No omega was used to calculate dN or dS
- Remember:

- $q_{IJ} = \begin{cases} 0 & \text{if more than one change} \\ \mu_{i_k j_k} h(S_{IJ}) & \text{if synonymous substitution} \\ \omega \mu_{i_k j_k} h(S_{IJ}) & \text{if non-synonymous substitution} \end{cases}$

# Conclusion

## Current and Next Steps

- Software currently exists, but it requires long computation
- We are running a MutSel model on PAML to understand various (input and intermediate) parameters and output

## Research Goals

- Compare Omega Models with Selection Coefficient
- Is one model better than the other?
- When is one model more appropriate? Under what conditions?